



Fisetin disposition and metabolism in mice: Identification of geraldol as an active metabolite

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ABSTRACT

Although the natural flavonoid fisetin (3,3',4',7-tetrahydroxyflavone) has been recently identified as an anticancer agent with antiangiogenic properties in mice, its *in vivo* pharmacokinetics and metabolism are presently not characterized. Our purpose was to determine the pharmacokinetics and metabolism of fisetin in mice and determine the biological activity of a detected fisetin metabolite. After fisetin administration of an efficacious dose of 223 mg/kg i.p. in mice, the maximum fisetin concentration reached 2.5 µg/ml at 15 min and the plasma concentration declined biphasically with a rapid half-life of 0.09 h and a terminal half-life of 3.1 h. Three metabolites were detected, one of which was a glucuronide of fisetin (M1), whereas another glucuronide (M2) was a glucuronide of a previously unknown fisetin metabolite (M3). HPLC–MS/MS analysis indicated that M3 was a methoxylated metabolite of fisetin (MW = 300 Da). The UV spectrum of M3 was identical to that of fisetin and standard 3,4',7-trihydroxy-3'-methoxyflavone (geraldol). In addition, because M3 co-eluted with standard geraldol in 4 different chromatographic ternary gradient conditions, M3 was therefore assigned to geraldol. Of interest, this metabolite was shown to achieve higher concentrations than fisetin in Lewis lung tumors. We also compared the cytotoxic and antiangiogenic activities of fisetin and geraldol *in vitro* and it was found that the latter was more cytotoxic than the parent compound toward tumor cells, and that it could also inhibit endothelial cells migration and proliferation. In conclusion, these results suggest that fisetin metabolism plays an important role in its *in vivo* anticancer activities.

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1. Introduction

Among the numerous products available from plants, the flavonoid superfamily plays a central role by their interesting pharmacological properties for several diseases, including cancer

prevention and therapy [1–4]. We, and others, have recently identified fisetin (3,3',4',7-tetrahydroxyflavone, Fig. 1), a flavonoid found in several fruits, vegetables, nuts and wine [5,6], as a lead compound possessing biological properties of potential interest in the prevention and/or treatment of cancer *in vivo* [7–9]. Fisetin possesses antioxidant and anti-inflammatory activities [10,11] and was found to be cytotoxic and antiangiogenic *in vitro* [12,13]. Fisetin is cytotoxic to various human cancer cell lines including leukemia (HL60) [14], breast (MCF7) [13], colon (HT29) [15], liver (SK-HEP-1, Caco-2) [15,16], neuroblastoma (SHEP, WAC-2) [13], prostate (LNCaP, PC3) [17], and also to endothelial cells [13]. Fisetin appears to interact with several molecular targets, including cyclin-dependent kinases [18–20], DNA topoisomerases I and II [21,22], urokinase [23], actin [24], and androgen receptor signaling [7]. It has also recently been found that fisetin induces a forced exit from mitosis by targeting the mitotic spindle checkpoint involving the inhibition of Aurora B activities required for the maintenance of normal spindle checkpoint signaling [25]. *In vivo*, fisetin has shown antiangiogenic activity after topical application in a rabbit model of corneal neovascularization [26].

Abbreviations: bFGF, basic fibroblast growth factor; DMEM, Dulbecco's modified Eagle's medium; DMSO, dimethylsulfoxide; EAhy 926, immortalized human umbilical vein endothelial cell line; HPLC, high-performance liquid chromatography; IC₅₀, inhibitory concentration for 50% of cells; LLC, Lewis lung carcinoma cells; MS/MS, tandem mass spectrometry; MTT, 1-(4,5-dimethylthiazol-2-yl)-3,5-diphenyltetrazolium; PBS, phosphate buffered saline.

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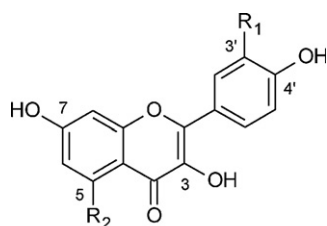


Fig. 1. Chemical structures of fisetin, geraldol and kaempferol. $R_1 = \text{OH}$, $R_2 = \text{H}$, fisetin or 3,3',4',7-tetrahydroxyflavone; $R_1 = \text{OCH}_3$, $R_2 = \text{H}$, geraldol or 3,4',7-trihydroxy-3'-methoxyflavone; $R_1 = \text{H}$, $R_2 = \text{OH}$, kaempferol or 3,4',5,7-tetrahydroxyflavone.

After systemic administration in mice, fisetin has shown interesting antitumor activity in several cancer models, including prostate, teratocarcinoma and lung carcinoma [7–9].

Although fisetin has shown activity *in vitro* and *in vivo* in mice, there is presently no information concerning the fisetin levels achieved in this preclinical species that could allow *in vitro* and *in vivo* comparisons. Moreover, the fisetin molecule bearing 4 hydroxyl substituents, including a catechol on the phenyl B ring, is therefore probably extensively metabolized *in vivo*. In the present study, at an effective dose of 223 mg/kg given by the i.p. route, we were therefore interested to determine the pharmacokinetics and metabolism of fisetin *in vivo* in mice.

We report here the pharmacokinetics of fisetin and its main metabolites in mice. In addition to glucuronide conjugates, we identified a 3'-methoxylated metabolite of fisetin (3,4',7-trihydroxy-3'-methoxyflavone or geraldol) as a major metabolite of fisetin. This methoxylated metabolite was found to be more cytotoxic than the parent compound and could also inhibit endothelial cells migration and proliferation.

2. Materials and methods

2.1. Chemicals and reagents

Fisetin (3,3',4',7-tetrahydroxyflavone), kaempferol (3,4',5,7-tetrahydroxyflavone), β -glucuronidase, dimethylsulfoxide (DMSO), and polyethylene glycol 200 (PEG200) were purchased from Sigma-Aldrich (Saint-Quentin Fallavier, France), and geraldol (3,4',7-trihydroxy-3'-methoxyflavone) was obtained from Extrasynthese (Genay, France). For HPLC studies, stock solutions of fisetin, kaempferol and geraldol were prepared in methanol and stored at -20°C protected from light. Methanol (HPLC grade) was purchased from Carlo Erba (Val de Reuil, France), glacial acetic acid was from Prolabo (Paris, France), and water was purified using a Milli-Q apparatus from Millipore Corporation. For cell culture, stock solutions of fisetin and geraldol were freshly prepared in DMSO and further diluted in cell culture medium. Matrigel and bFGF were obtained from BD Biosciences (Le Pont de Claix, France). All other chemicals were obtained from commercial sources and were of the highest purity available.

2.2. Fisetin pharmacokinetics in mice

Female 8 weeks old C57BL/6J mice (mean body weight 18–22 g) were purchased from Janvier (Le Genest-St-Isle, France). After an overnight fasting period, mice were injected intraperitoneally (i.p.) with fisetin (223 mg/kg body weight) dissolved in 100 μl of PEG200/DMSO (7:3, v/v). This dose level was chosen because it was nontoxic (based on body weights) when administered i.p. (daily $\times 5$) for 2 consecutive weeks and also because this dose was found active in Lewis lung carcinoma bearing mice [9]. Blood samples ($n = 3$ per sampling time) were collected by cardiac puncture with

heparinized syringes after sacrificing the animals by CO_2 at the following times: pre-administration (time 0), and at 0.25, 0.5, 1, 1.5, 2, 4, 8, 15 and 24 h post-dosing. Blood samples were transferred into polypropylene tubes and centrifuged at $12,000 \times g$ for 10 min to obtain the plasma which was kept at -20°C until HPLC analysis. For the determination of fisetin and metabolites in tumors, pieces of Lewis lung carcinoma (LLC) were injected subcutaneously into the right flank of female C57BL/6J mice (5 mm^3) on day 0. Fifteen days later, when the tumors reached a volume of 500–1000 mm^3 , mice were injected with fisetin (223 mg/kg, i.p.) dissolved in 100 μl of polyethylene glycol 200/DMSO (7:3, v/v), and 15 min later, the mice were sacrificed and the tumors were harvested, rinsed with ice-cold PBS and transferred into a polypropylene tube and kept at -20°C until analysis. All animal experiments have been carried out in accordance with institutional and French regulations concerning the protection of animals used for experimental and other scientific purposes, and with the European Commission regulations.

2.3. Sample preparation and extraction

Plasma samples, tumor or normal tissues homogenates (prepared with 1 ml of PBS using a Teflon potter at a speed of 50 rpm with 3 strokes), were extracted by mixing (Vortex mixer, 1 min) with a 5-fold volume of cold methanol containing kaempferol (3,4',5,7-tetrahydroxyflavone, Fig. 1), as an internal standard (final concentration 3.3 $\mu\text{g}/\text{ml}$). The mixture was put on ice for one hour to precipitate the proteins, and the samples were centrifuged at $12,000 \times g$ for 10 min at 4°C . The upper phase was transferred to another polypropylene tube and 100 μl was injected onto a Shimadzu HPLC-UV system described below.

2.4. Standard curves and extraction

Stock solutions of fisetin, geraldol and kaempferol (cf., Fig. 1) were prepared in methanol at 100 $\mu\text{g}/\text{ml}$ and stored at -20°C in the dark. Working standard solutions of increasing concentrations of fisetin and geraldol were prepared on the day of analysis by diluting the stock solution. The standard curves of fisetin and geraldol in mouse plasma, tumors, or normal tissues were built using the ratio of the area of fisetin or geraldol over the area of the internal standard (kaempferol) and were linear within the concentration range used (0.01–20 $\mu\text{g}/\text{ml}$) with correlation coefficient > 0.99 . The percent extraction from plasma, tumors, or normal tissues was 91% and the lower limit of quantification was 0.01 $\mu\text{g}/\text{ml}$. The apparent concentration of the metabolites was calculated based on the fisetin standard curve.

2.5. HPLC system for pharmacokinetic studies

For pharmacokinetic studies we used an HPLC-UV system (Shimadzu CLASS-VP[®], version 5.3), equipped with a C_{18} column (Beckman Ultrasphere ODS, 5 μm ; 4.6 mm \times 25 mm) thermostated at 20°C , and a UV detector set at 360 nm. The mobile phase was composed of 52% methanol and 48% aqueous acetic acid (2%), at a flow rate of 1 ml/min.

2.6. Pharmacokinetic parameters determination

Fisetin plasma concentrations were analyzed using noncompartmental analysis and also using a bicompartmental model with extravascular administration. The following noncompartmental pharmacokinetic parameters were derived using standard methods [27]: area under the plasma concentration versus time curve from time zero to the time of the last measurable concentration (AUC_{0-t}) calculated by the trapezoidal method; maximum

concentration (C_{\max}); mean residence time (MRT); terminal half-life; apparent intraperitoneal clearance (CL/F) and volume of distribution at steady state (V_{ss}/F). Pharmacokinetics parameters were also determined by fitting the fisetin plasma concentration versus time to a two-compartmental model with extravascular administration using the Graph Pad Prism software.

2.7. Identification of metabolites

2.7.1. Hydrolysis of glucuronide conjugates

To hydrolyze glucuronide conjugates, plasma or tumor homogenate samples were adjusted to pH 5.0 with an equal volume of 50 mM acetate buffer, and incubated at 37 °C for 2 h in the presence of 1000 units of β -glucuronidase (Sigma No. G-0751). The hydrolyzed samples were submitted to the above described protocol of methanolic extraction and injected onto the HPLC system.

2.7.2. Identification of metabolite M3 using UV and mass spectrometry

Metabolite M3 was analyzed online using an HPLC–MS/MS system composed of an HPLC apparatus (Surveyor[®], ThermoElectron) equipped with a UV diode array detector and coupled with a mass spectrometer (LCQ-Advantage[®], ThermoElectron). A C₁₈ reversed-phase column (EC. 250/2 Nucleodur Sphinx RP, 5 μ m, 250 mm \times 2.1 mm, Macherey-Nagel) thermostated at 30 °C was used. The metabolites (injection volume: 15 μ l) were resolved with the following ternary gradient using methanol (A), acetonitrile (B) and aqueous 0.1% TFA (C) at a flow rate of 0.2 ml/min: from 0 to 50 min, 7–35% A, 4–19% B, 89–46% C; from 50 to 60 min, 35% A, 19% B, 6% C (gradient 1). The mass spectrometer consisted of an electrospray ionization system used in positive ion polarity mode and ion trap mass analyzer. The operating parameters were as follows: the spray needle voltage was set at 4.0 kV and the spray was stabilized with a nitrogen sheath gas. Electrospray capillary voltage was 4.0 V, nitrogen was used as auxiliary gas, and the capillary temperature was 250 °C. The Xcalibur[®] software was used for data acquisition and analysis.

2.7.3. Identification of M3 using retention times in various gradient systems

To ascertain the identity of M3, this metabolite and authentic geraldol were separately injected using the HPLC system with gradient 1 described in Section 2.7.2, and three other ternary gradients composed of methanol (A), acetonitrile (B) and aqueous 0.1% TFA (C) (flow rate 0.2 ml/min) as follows: gradient 2 – from 0 to 70 min, 7–35% A, 4–19% B, 89–46% C; gradient 3 – from 0 to 50 min, 15–35% A, 10–19% B, 75–46% C; gradient 4 – from 0 to 70 min, 15–35% A, 10–19% B, 75–46% C. For each gradient, the logarithm of the retention factor ($\log k$) of M3 and geraldol was calculated using retention time and dead time. The retention factor relates to retention time, but is preferred because it is dimensionless and independent of unexpected variations in mobile phase flow. In contrast to retention time, the $\log k$ can unequivocally be ascribed to structural changes, as previously used for flavonoids by Stefova et al. [28]. To further ascertain the identity of M3, we also injected an equimolar mixture of geraldol and M3 using all 4 gradient systems.

2.8. In vitro cytotoxic activity of fisetin and metabolite

The murine Lewis lung carcinoma (LLC) cell line, the normal NIH 3T3 murine fibroblast cell line and the EAhy 926 endothelial cell line were grown in Dulbecco's modified Eagle's medium (DMEM) containing 2 mM L-glutamine, 10% fetal bovine serum, 100 U/ml penicillin and 100 μ g/ml streptomycin (37 °C, 5% CO₂).

Exponentially growing cells were plated onto 96-well plates at 5000 cells per well in 200 μ l. Twenty-four hours later, cells were exposed for 24 or 48 h to fisetin or geraldol at increasing concentrations (0–1 mM). Viability was assessed using the MTT test and absorbance was read at 562 nm in a microplate reader (BioKinetics Reader, EL340). Appropriate controls with DMEM only and MTT were run to subtract background absorbance. Experiments were run in quadruplicate and repeated 3 times. Control cells were exposed to 1% DMSO which was not cytotoxic. The results are presented as the inhibitory concentrations for 50% of cells (IC₅₀) for a 24 or 48 h exposure time relative to controls.

2.9. Cell migration assay (scratch wound assay)

EAhy 926 endothelial cells were grown to confluence and a wound was introduced by clearing an area of the monolayer using a 100 μ l pipet tip. After wounding, digital photographs were recorded of wounded areas of each well at a magnification of $\times 100$ (time 0 h). Medium was changed, and bFGF (10 ng/ml), fisetin or geraldol was added at increasing concentrations (0, 22, 44 and 88 μ M) to the media containing 2.5% of FBS. After a 24 h incubation period, digital photographs were recorded of the cleared areas of each well. Migration was evaluated by manually drawing the distance of the wound area (d) at T_{0h} and T_{24h} time on a computer screen and distance's values were obtained using the ImageJ software [29]. The results were expressed as a percent of the controls using the following formula: $100 \times [1 - (dT_{0h} - dT_{24h} \text{ of treated cells}) / (dT_{0h} - dT_{24h} \text{ of control cells})]$. Experiments were performed in triplicate for each concentration and were repeated 3 times.

2.10. Formation of capillary-like structures

Analysis of capillary-like structure formation was performed using EAhy 926 endothelial cells grown on Matrigel. Fifty microliter of the Matrigel solution was applied to each well on a 96-well plate and incubated for 30 min at 37 °C. EAhy 926 cells (1×10^4) were suspended in 100 μ l of medium, plated onto the gel matrix and incubated at 37 °C. Adherent cells received bFGF (10 ng/ml) and fisetin or geraldol at increasing concentrations (0, 22, 44 and 88 μ M). After a 24 h exposure time, *in vitro* angiogenesis was assessed by counting the capillary-like structures in each well at $\times 100$ magnification under a light microscope (Zeiss). The *in vitro* anti-angiogenic effect was calculated by the following formula: $100 \times [1 - (\text{number of capillary-like tube of treated cells}) / (\text{number of capillary-like tube of control cells})]$. Experiments were performed in duplicate for each condition and repeated 3 times.

2.11. Statistical analysis

Results are expressed as the mean \pm SEM of at least 3 independent experiments. Statistical differences between means were assessed using Student's *t* test for unpaired data. If unequal variance was observed, the Welch's correction was applied. A *P* value ≤ 0.05 was considered significant.

3. Results

3.1. Plasma and tissue pharmacokinetics of fisetin and its metabolites

A representative chromatogram depicting the separation of fisetin and its metabolites in mouse plasma is presented in Fig. 2. In these chromatographic conditions (reversed-phase mode), fisetin eluted at 40 min and the internal standard (kaempferol) at 53 min. Fisetin plasma pharmacokinetics after an i.p. administration at a dose of 223 mg/kg is presented in Fig. 3 and Table 1. The maximum

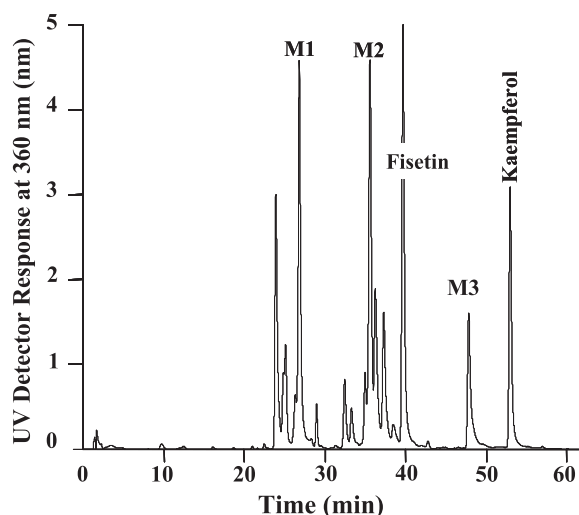


Fig. 2. Representative HPLC chromatogram showing the separation of fisetin and its main metabolites formed *in vivo* (M1, M2, M3) 15 min after the intraperitoneal injection of fisetin at a dose of 223 mg/kg. Kaempferol was used as an internal standard. In control plasma, there was no detectable peak present after the first 5 min.

plasma concentration reached 2.53 $\mu\text{g/ml}$ at 15 min post dosing and declined thereafter with a first rapid alpha half-life of 0.09 h (5.4 min), followed by a longer terminal disposition half-life beta of 3.12 h. The total area under the plasma versus time curve (AUC) was 4.0 ($\mu\text{g/ml}$) \times h, the apparent intraperitoneal clearance (CL/F) was 1.12 L/h, and the Vss/F was 5.1 L.

As shown in Fig. 2, in addition to the fisetin peak, several other peaks not present in control plasma were also detected. Of those peaks, two metabolites eluted before fisetin (M1 at 26 min and M2 at 35 min), whereas another metabolite (M3) eluted after fisetin with a retention time of 48 min. Fig. 4 presents the relative concentrations of these metabolites as a function of time using fisetin as a reference. It can be observed that metabolites M1 and M2 could achieve apparent higher concentrations than the parent compound, whereas metabolite M3 apparent concentrations were lower compared to fisetin. Pharmacokinetic parameters of the metabolites are presented in Table 1.

To obtain some information on fisetin and metabolites levels in normal tissues, mice were sacrificed 30 min after i.p. injection of fisetin and liver, intestines, kidneys, lungs and spleen were

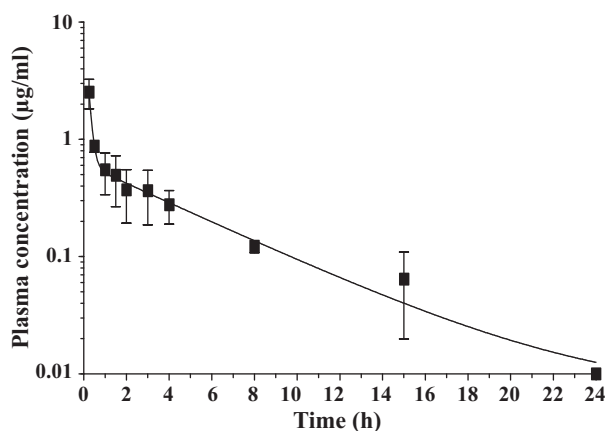


Fig. 3. Pharmacokinetic profile of fisetin in mouse plasma. After an intraperitoneal injection of 223 mg/kg, fisetin concentration was determined by HPLC as described in Section 2. Mean \pm SEM of 3 mice per time point. Solid line represents the fitting to an extravascular 2 compartment model.

Table 1

Fisetin and metabolites pharmacokinetic parameters.

	C_{\max} ($\mu\text{g/ml}$)	AUC ($\mu\text{g/ml}$) \times h	MRT (h)	Half-lives (h)
Fisetin	2.53	4.00	4.51	3.12
Metabolite M1	6.54	40.71	11.27	7.81
Metabolite M2	2.22	13.63	9.95	6.89
Metabolite M3	0.07	0.17	0.65	0.45

C_{\max} , maximum plasma concentration; AUC, area under the plasma concentration versus time curve from time zero to the time of the last measurable concentration; MRT, mean residence time.

harvested. Fisetin and metabolites levels presented in Table 2 show that fisetin concentrations were highest in liver, intestines and kidneys. Metabolite M1 highest concentrations were mainly found in plasma and kidneys. For metabolites M2 and M3, the highest concentrations were in kidneys and intestines.

We next submitted plasma samples to β -glucuronidase digestion, and the M1 and M2 metabolite peaks were completely abolished, whereas the fisetin and the M3 peaks were markedly increased as shown in Fig. 5 (compare with Fig. 2 without β -glucuronidase). This observation indicated that M1 and M2 were glucuronides of fisetin and metabolite M3, respectively.

3.2. Identification of metabolite M3

To identify metabolite M3, the peak at 48 min was subjected on line to mass spectrometry analysis and UV spectrum determination. The mass spectrum of M3 presented a molecular ion m/z value of 301 Da $[M+H]^+$, thus corresponding to a molecular weight of 300 Da (Fig. 6A). In the same conditions, fisetin presented a $M+H$ m/z value of 287 Da corresponding to a molecular weight of 286 Da (data not shown). Therefore, the gain of 14 atomic mass units observed with M3 over the parent compound corresponded to the addition of a CH_2 on the fisetin molecule, indicating that metabolite M3 was a methylated compound of fisetin. However, because fisetin possesses 4 hydroxyl groups, the exact localization of the methoxyl on the fisetin molecule could not easily be identified.

In addition, the UV spectrum of M3 was found superimposable to that of fisetin with identical maxima at 245 nm, 320 nm (shoulder) and 360 nm (Fig. 6B). Among the possible methylated

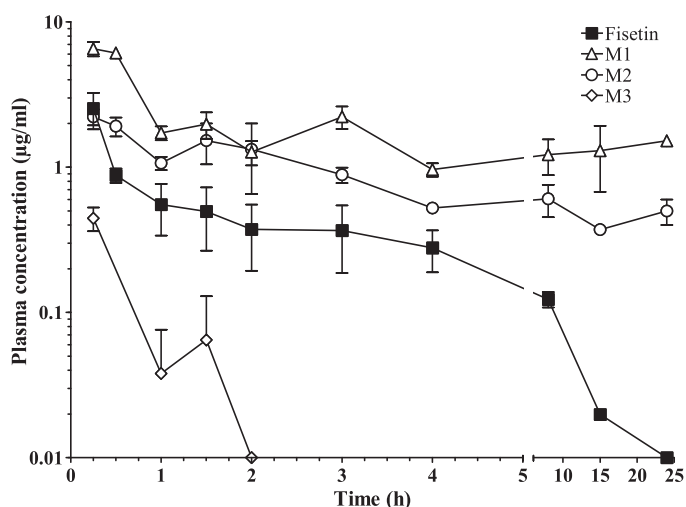


Fig. 4. Plasma concentration of the main fisetin metabolites (M1, M2 and M3) as a function of time after the intraperitoneal injection of fisetin (223 mg/kg). Fisetin and metabolites concentrations were determined by HPLC as described in Section 2. The apparent concentration of the metabolites was calculated based on the fisetin standard curve. Mean \pm SEM of 3 mice per time point. Solid squares, fisetin; triangles, metabolite M1; circles, metabolite M2; diamonds, metabolite M3.

Table 2

Fisetin and its metabolites distribution in plasma and normal tissues in mice.

Tissues	Concentration ($\mu\text{g}/\text{ml}$ or $\mu\text{g}/\text{g}$ tissue)			
	Fisetin	M1	M2	M3
Plasma	1.14 ± 0.12	6.12 ± 0.47	1.93 ± 0.29	0.10 ± 0.01
Liver	39.15 ± 12.85	0.66 ± 0.22	2.02 ± 1.31	2.23 ± 0.38
Intestines	80.11 ± 33.82	0.12 ± 0.08	4.17 ± 3.24	5.05 ± 0.49
Kidneys	83.69 ± 10.45	4.23 ± 1.88	49.03 ± 22.65	14.25 ± 6.54
Lungs	8.00 ± 3.31	0.69 ± 0.46	5.67 ± 5.69	1.57 ± 0.67
Spleen	2.00 ± 0.50	1.03 ± 1.37	1.30 ± 1.01	1.71 ± 1.67

Mice were administered fisetin intraperitoneally at a dose of 223 mg/kg and tissues were harvested 30 min later for HPLC analyses, as detailed in Section 2. Values are the mean \pm SEM of 3 independent determinations.

fisetin candidates, the 3,4',7-trihydroxy-3'-methoxyflavone or geraldol (cf., Fig. 1), therefore appeared as a possible candidate. Geraldol UV spectrum was indeed found superimposable to that of M3 and fisetin (Fig. 6B). Also the mass spectrum of authentic geraldol was also superimposable to the mass spectrum of M3 (data not shown).

Authentic geraldol was also tested for its HPLC retention time and it was found to co-elute with metabolite M3 in 4 different ternary elution systems (Table 3), thus further substantiating that M3 was indeed geraldol. In addition, the logarithm of the retention factor ($\log k$) of M3 and geraldol yielded the same values, further substantiating the identity of M3 as geraldol. Considering M3 molecular mass identical to geraldol, its UV spectrum superimposable to both fisetin and geraldol spectra, its retention times and $\log k$ identical to authentic geraldol in 4 ternary gradient systems allowed us to ascribe M3 as being geraldol.

3.3. Tumor concentration of fisetin and its metabolites

At 15 min after the administration of fisetin i.p. (223 mg/kg) to tumor bearing mice (Lewis lung carcinoma), the plasma and tumor concentrations of fisetin and its main metabolites were also determined by HPLC. Glucuronides M1 and M2 presented higher concentrations in the plasma compared to tumors (compare Fig. 7A and B). However, it was of interest to note that geraldol was the metabolite that reached the highest concentrations in tumors (Fig. 7B). Indeed, geraldol concentrations in tumors were 4.3 times higher than the fisetin concentration in this tissue at this sampling

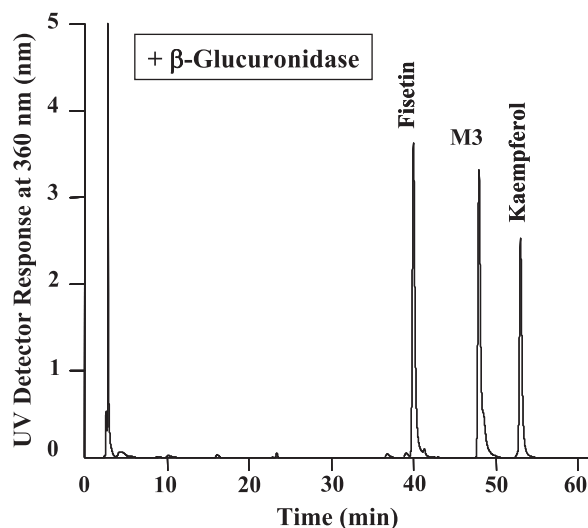


Fig. 5. Representative HPLC chromatogram showing the β -glucuronidase digestion of metabolites M1 and M2 to yield fisetin and metabolite M3. Mice received an intraperitoneal injection of fisetin at a dose of 223 mg/kg. Kaempferol was used as an internal standard.

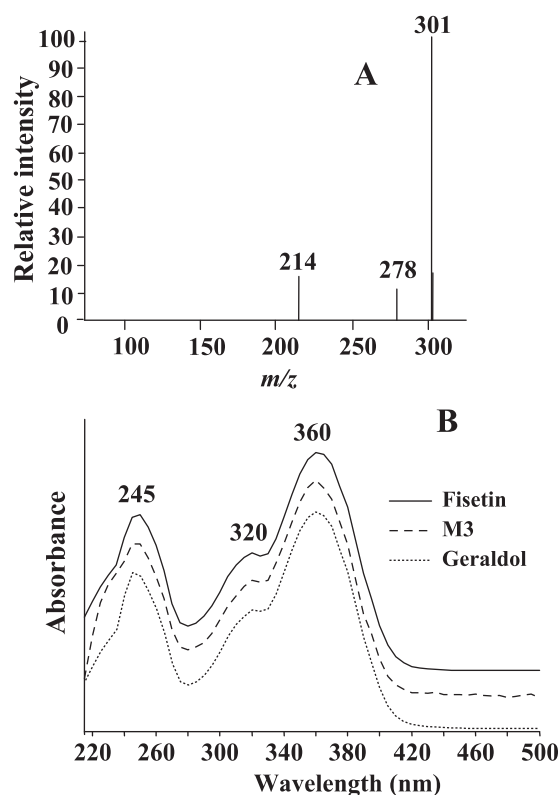


Fig. 6. Mass and UV spectra of metabolite M3. (A) Representative mass spectrum of metabolite M3 obtained on line with an HPLC–MS/MS equipment detailed in Section 2. (B) UV spectra of fisetin, metabolite M3, and geraldol obtained online using a diode array detector.

time. Considering this observation, we were next interested to investigate further the geraldol cytotoxic and antiangiogenic properties.

3.4. Cytotoxic activity of fisetin and geraldol on Lewis lung carcinoma (LLC) cells, endothelial cells (EAhy 926), and normal cells (NIH 3T3)

The effect of fisetin and metabolite geraldol on the viability of Lewis carcinoma cells, endothelial cells (EAhy 926), and normal NIH 3T3 cells were also tested (Fig. 8). A dose-dependent decrease in viability of the three cell lines after a 24 h exposure time to both flavonoids could be observed. The most cytotoxic flavonoid on LLC cells was found to be geraldol with an IC_{50} (the concentration producing 50% reduction of viability) of 24 μM , about 2.5-fold lower than that of fisetin (59 μM) (Fig. 8A). Concerning the effect on EAhy 926 endothelial cells viability, fisetin and geraldol showed similar IC_{50} (76 and 72 μM , respectively) (Fig. 8B). However, it was

Table 3

Retention times of fisetin and geraldol in various HPLC ternary gradient systems.

Gradient system ^a	Retention times in minutes ($\log k$) ^b		
	Fisetin	Geraldol	Metabolite M3
1	40.0 (0.561)	47.7 (0.590)	47.7 (0.591)
2	49.4 (0.596)	60.1 (0.628)	60.1 (0.628)
3	28.6 (0.504)	39.0 (0.557)	39.0 (0.557)
4	32.8 (0.528)	46.6 (0.586)	46.6 (0.586)

^a The ternary gradient systems used were composed of methanol (A), acetonitrile (B) and water containing 0.1% TFA, as follows: gradient 1 – from 0 to 50 min: 7–35% A, 4–19% B, and 89–46% C; gradient 2 – from 0 to 70 min: 7–35% A, 4–19% B, 89–46% C; gradient 3 – from 0 to 50 min: 15–35% A, 10–19% B, 75–46% C; gradient 4 – from 0 to 70 min: 15–35% A, 10–19% B, 75–46% C. The reversed-phase HPLC system used is described in Section 2.

^b The retention factor k was calculated as in Stefova et al. [28].

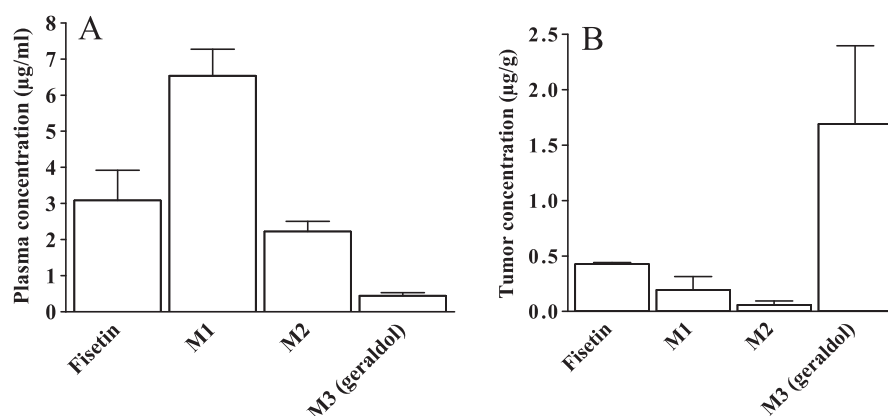


Fig. 7. (A) Plasma and (B) tumor concentrations of fisetin and its main metabolites M1, M2 and M3 (geraldol), 15 min after the i.p. administration of fisetin (223 mg/kg). M1 and M2 are glucuronides of fisetin and geraldol, respectively, whereas M3 is geraldol. Concentrations were determined by HPLC as described in Section 2.

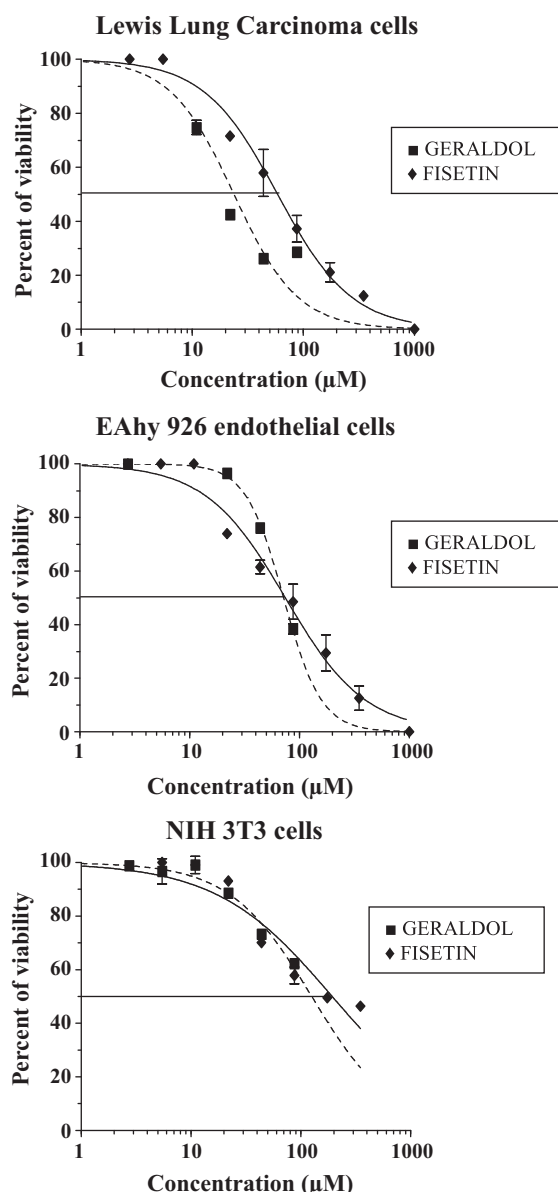


Fig. 8. Comparative cytotoxicity of fisetin and geraldol on various cell lines.

of interest that the normal NIH 3T3 cells were significantly less sensitive to fisetin and geraldol than either LLC or endothelial cells, with IC₅₀ of 195 µM and 128 µM, respectively (Fig. 8C).

3.5. Fisetin and geraldol effect on endothelial cells migration in vitro

We next compared the effect of various concentrations (22, 44, and 88 µM) of fisetin and its metabolite geraldol on EAhy 926 endothelial cells migration using the scrape wound assay. Fig. 9 illustrates that at 24 h post-wounding of confluent EAhy 926 endothelial cells, the control bFGF (10 ng/ml) treated cells migrated and totally filled the scraped area, as expected. Although fisetin and geraldol could inhibit significantly endothelial cells migration at 44 µM (Fig. 9), fisetin was found to be more active than geraldol at this concentration, and also at the lower concentration of 22 µM (Fig. 10). Although the inhibition of endothelial cells migration was dose-dependent for both flavonoids, the calculated IC₅₀ for the antimigration effect showed that fisetin had a significantly ($P < 0.001$) stronger antimigration effect compared to geraldol (43 ± 0.8 µM versus 64 ± 0.8 µM, respectively) (Fig. 10).

3.6. Fisetin and geraldol effect on the capillary-like structure formation on Matrigel

To examine whether geraldol could directly inhibit angiogenesis, its effect on bFGF-mediated tube formation by endothelial cells was investigated at concentrations ranging from 3 to 300 µM. As expected, EAhy 926 endothelial cells plated on Matrigel with bFGF (10 ng/ml) formed a capillary-like network within 24 h (data not shown). Fisetin could prevent the formation of the capillary-like network in a dose-dependent fashion with an IC₅₀ of 51 ± 7 µM, whereas no significant antiangiogenic effect of geraldol could be observed at the highest achievable *in vitro* concentration of 88 µM, because geraldol was not soluble at higher concentrations (data not shown).

4. Discussion

The natural flavonoids are attracting interest in cancer prevention and therapy because these compounds are relatively nontoxic compared to other chemotherapeutic agents used in cancer therapy [1–4]. Although the dietary flavonoid fisetin has recently been shown to be an interesting anticancer active agent in mice [7–9] and to possess antiangiogenic properties *in vitro* [12,13] and *in vivo* [9], the blood levels achieved by this compound after

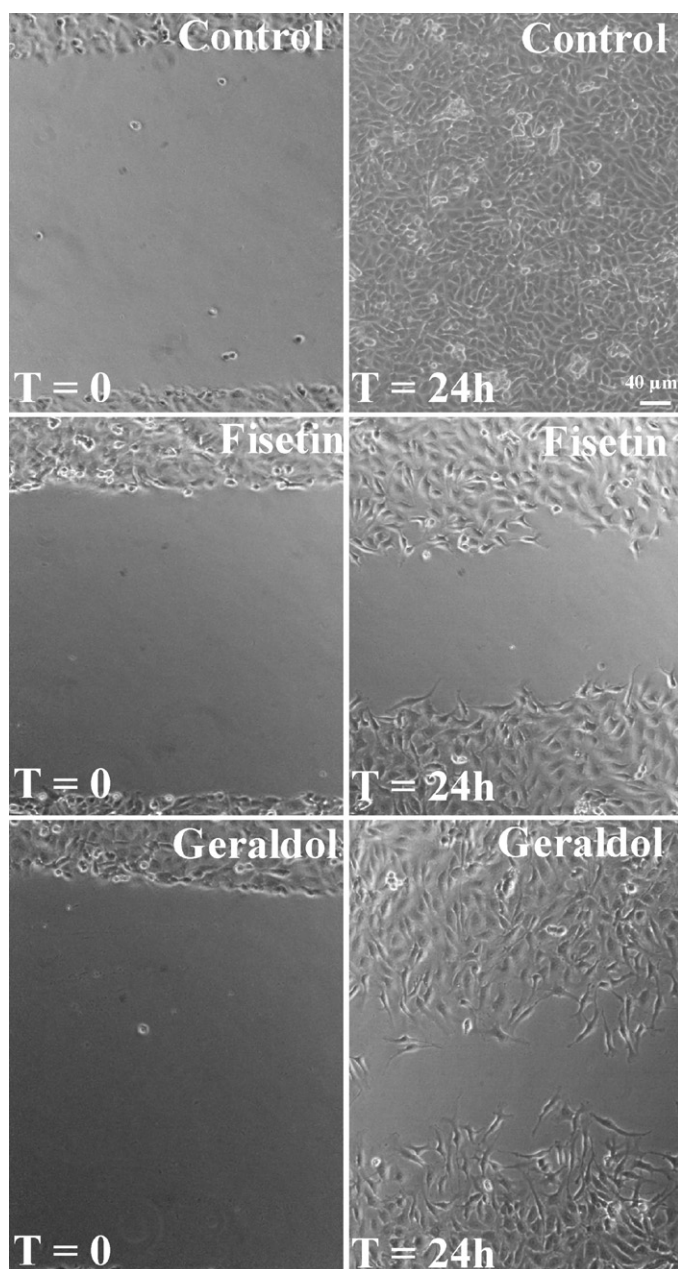


Fig. 9. Effect of fisetin and its metabolite geraldol on endothelial cell migration (scratch wound assay). Confluent EAhy 926 endothelial cells were grown to confluence and an area was cleared using a 100 μ l pipet tip. Digital photographs were recorded at a magnification of $\times 100$ at time 0 and 24 h later after the addition of bFGF (10 ng/ml) for the control, or with bFGF plus 44 μ M of either fisetin or geraldol. Scale bar, 40 μ m.

systemic administration and its metabolism have not previously been investigated in mice. The purpose of this study was therefore to study fisetin pharmacokinetics in mice, to determine fisetin principal metabolic routes and to evaluate the biological activity of a detected metabolite.

Concerning the pharmacokinetics of fisetin in mouse plasma, we found that it could reach a concentration of 2.5 μ g/ml at 15 min and that it displayed a relatively long terminal half-life in plasma of 3.12 h after an i.p. administration of 223 mg/kg. Although the plasma concentrations are relatively low, it is possible that the long residence time could be responsible for the remarkable *in vivo* antitumor effect in mice at this dose level [9]. Nonlinear pharmacokinetics may also be involved in the fisetin long residence time at this dose level, as nonlinear pharmacokinetics

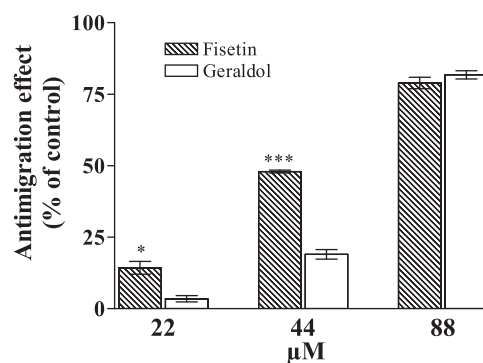


Fig. 10. Comparative antimigration effects of fisetin and its metabolite geraldol. Endothelial EAhy 926 cells were grown to confluence and cell wounding was induced using a pipet tip, as described in Section 2. After a 24 h exposure time at the indicated concentrations of fisetin (hatched bars) and geraldol (open bars) in presence of bFGF, the relative antimigration effect was calculated as described in Section 2. Mean \pm SEM of 3 independent experiments. Asterisks indicate a statistically significant difference relative to controls at the $P < 0.05$ (*) or $P < 0.001$ (***) level (Student's *t* test).

has already been reported for other flavonoids [30]. Potential accumulation of fisetin and/or metabolites in tumor tissue could also play a role in the fisetin antitumor activity. Compared to fisetin pharmacokinetics in rat in which negligible levels of free fisetin and a short half-life of 2.7 min were reported [31], it thus appears that the mouse is eliminating fisetin at a much slower rate (half-life = 3.12 h), probably due to a lower conjugation capacity through glucuronidation in mice. Indeed, such significant species differences in clearance have also been observed for the flavonoid analogue 5,6-dimethylxanthone-4-acetic acid (DMXAA), which is also mainly metabolized through glucuronidation, where *in vitro* intrinsic clearance in mice was found to be 15-fold lower than that in rats, and *in vivo* plasma clearance was 1.8-fold smaller in mice compared to rat plasma clearance [32].

We also observed that metabolites M1 and M2, which were identified as glucuronides, displayed longer terminal half-lives compared to fisetin. This appears unusual since these two metabolites are glucuronides of fisetin and M3, and shorter half-lives could have been expected. However such a pharmacokinetic behaviour has also been reported for fisetin in rats where a half-life of 2.7 min was found for the aglycone, whereas the glucuronides were showing a half-life of 40.7 min [31]. Such a longer half-life for genistein sulfates/glucuronides has also been observed in rats compared to the parent aglycone [33].

In this study, we have also identified a methylated metabolite of fisetin which was assigned to the 3,4',7-trihydroxy-3'-methoxyflavone (geraldol). This metabolite, reported here for the first time to our knowledge, is probably generated through catechol-*O*-methyl transferase (COMT) metabolism because fisetin possesses a catechol function on the B ring. COMT catalyzes the transfer of a methyl group from *S*-adenosyl-L-methionine to polyphenols possessing an *o*-diphenolic (catechol) moiety. COMT can be found as membrane bound and soluble forms and is expressed in brain and most peripheral tissues including liver, kidneys and small intestine in the mouse, although there are large differences between species [34,35]. Methylation has already been reported for other flavonoids possessing a catechol function, e.g., quercetin, catechin, caffeic acid, and luteolin (reviewed in [35]). Methylation of flavonoids generally occurs predominantly in the 3'-position of the polyphenol, but a minor proportion of 4'-*O*-methylated product can also be formed.

Although our mass spectrometry data indicated the presence of a methoxyl group on the fisetin molecule, the exact position of this substituent could not be determined by this technique alone. The formal identification of the methoxylated metabolite of fisetin was

therefore further investigated using UV spectroscopy which is particularly useful for the identification of flavonoids [36]. It is known that methylation of hydroxyl groups in positions 3, 5, 7, or 4' produces hypsochromic shifts (shorter wavelengths) of the characteristic absorption bands in the UV-spectra of flavones, but does not significantly affect the shape of the UV-spectra when it occurs in other positions [37]. In our case, metabolite M3 did not present a hypsochromic shift compared to the parent compound and their UV spectra were superimposable. Therefore, the methylation of the 3, 4', and the 7 positions could therefore be ruled out as possible methylated positions of fisetin.

We further ascertained the identity of metabolite M3 as being the 3,4',7-trihydroxy-3'-methoxyflavone or geraldol using the log *k* value obtained by reversed-phase HPLC. It was already demonstrated that log *k* values are significantly different for the same flavone if methylated in the 3' or the 4' position [28]. Using the same approach, the log *k* values obtained with 4 different ternary mobile phases were strictly identical for geraldol and metabolite M3, therefore substantiating further that M3 was indeed the 3'-methyl-fisetin, and could not be the 4'-methyl-fisetin. Concerning the possible methylation on position 3 of fisetin, this would have resulted in a decrease in the retention time in the reversed-phase HPLC mode, owing to its inductive effect which increases the influence of the polar carbonyl group [28]. Methylation of the hydroxyl group in position 3 results in a lack of hydrogen bonding between 3-OH and the carbonyl group (at C4) which results in an increased polarity of the molecule and consequently a decrease in retention time. In contrast, an increase in retention time was observed compared to fisetin, which further rules out a methylation at the 3 position of fisetin. Considering metabolite M3 molecular mass identical to geraldol, its UV spectrum identical to fisetin and geraldol, and identical retention times to geraldol in 4 different ternary gradient systems, the identity of M3 was therefore assigned to geraldol.

Of particular interest, it was also observed that this 3'-methoxylated metabolite of fisetin could achieve higher concentrations in tumors compared to fisetin, an observation probably linked to the fisetin *in vivo* remarkable anticancer activity already noted by some investigators [7–9]. This high tumor geraldol levels could result from either an increased *in situ* formation or this metabolite, or from hydrolysis of its glucuronide that could regenerate the aglycone. Indeed, a high COMT activity in the tumor is possible because an increased expression of this enzyme has recently been observed in cancer [38]. Concerning the possible regeneration of the aglycone from its corresponding glucuronide, it has been reported that Lewis lung tumors express a high β -glucuronidase activity that could hydrolyze the glucuronide to its geraldol aglycone [39].

From a metabolic standpoint, the methylation of fisetin *in vivo* could be viewed as a favorable metabolic route, because methylation usually results in a higher metabolic stability of flavonoids that could favor its retention *in vivo* [40]. Because geraldol can be glucuronidated, as shown in this work, and could therefore be secreted in bile, bacterial glucuronidase present in the intestines could hydrolyze this metabolite and give back the aglycone (geraldol) that could be absorbed from the intestines, therefore possibly contributing to a longer residence time *in vivo* for this metabolite. In addition, the presence of a methoxy group on the A or B ring of the flavonoid also seems to protect the structure from bacterial degradation in faeces, therefore probably contributing to longer exposure times *in vivo* by allowing the reabsorption from the intestines of the intact methylated metabolite [41].

However, we observed in this work that the geraldol formed *in vivo* is rapidly cleared from the plasma as shown by its short half-life, although this metabolite is less polar and probably more metabolically stable than fisetin. We have no obvious explanation

for this observation, except that upon geraldol formation *in vivo*, this metabolite could be rapidly metabolized to other metabolites (e.g., glucuronides and/or sulfates), which could perhaps explain its rapid clearance.

The fisetin antiproliferative/cytotoxic activity determined in this study on Lewis lung carcinoma cells is in line with its cytotoxic activity reported on other cancer cell lines, e.g., in prostate [17], liver [16], colon [18], and leukemia cell lines [14]. With regard to the comparison of the biological activity of fisetin and its metabolite geraldol, it was of interest to note that the metabolite was more cytotoxic than the parent compound on Lewis lung carcinoma cells showing a 2.5-fold lower IC₅₀ value. This increased cytotoxicity of the metabolite associated with the higher concentrations of geraldol in tumor could give fisetin a selective anticancer activity advantage *in vivo*. Also of interest, both fisetin and geraldol were found several folds less cytotoxic toward normal NIH 3T3 cells compared to tumor cells, and this could give these compounds an important *in vivo* advantage in terms of therapeutic index, because they would be less toxic to normal cells compared to cancer cells. This relative nontoxicity of flavonoids to normal cells was also noted for other normal cell lines [13]. This tumor cell selectivity was also observed on prostate cancer cells that were shown to be more vulnerable to fisetin compared to normal prostate cells [17]. This relatively uncommon cancer cell selectivity could therefore confer fisetin a valuable advantage for *in vivo* anticancer treatment.

In addition, it should be mentioned that because of the presence of 4 hydroxyl substituents on the fisetin molecule, several glucuronide conjugates were also detected at high concentrations in mouse plasma in the present mouse study, and also in rat plasma [31]. These metabolites could also play a role in the overall antiangiogenic effects observed *in vivo* in mice, because some flavonoid sulfates and/or glucuronides have been reported to display bioactivities sometimes superior to their aglycones (free forms) [42–45]. Future studies aimed at determining potential high partition of metabolites and/or accumulation in tumor tissue should also be considered, as well as a more complete pharmacokinetic profile of fisetin and its metabolites in tumors should also be assessed.

The interesting bioactivity of geraldol shown in the present work has also to be put in perspective of a recently published work on quercetin (3',3,4',5,7-pentahydroxyflavone), a close parent of fisetin bearing an additional 5-hydroxyl substituent. A 3'-O-methylated metabolite of quercetin, i.e., isorhamnetin, was shown to possess potent inhibitory activity of cyclo-oxygenase-2 (COX-2), an enzyme mechanistically linked to carcinogenesis [46]. Also in line with this observation, quercetin metabolites have been found to possess antioxidant properties [47–49], and to inhibit xanthine oxidase and lipoxygenase [50].

Regarding the effect of fisetin and geraldol on the formation of capillary-like structures, the parent compound fisetin has previously been shown to be active in this model [9], whereas geraldol could not achieve an active concentration in this model probably due to its inherently low water solubility. It is however possible that longer exposure time to geraldol (>24 h) could nonetheless participate to an antiangiogenic response *in vitro*. Considering the above results, it is therefore possible that fisetin antitumor action *in vivo* could involve a dual mechanism of action, i.e., the parent compound fisetin would exert its strong antiangiogenic action directly, whereas its main metabolite geraldol would then exert its strong cytotoxicity against tumor cells.

In conclusion, we have determined fisetin pharmacokinetics in mice, for the first time to our knowledge, and have also identified geraldol, a previously unknown metabolite. In addition, we have shown that the metabolite geraldol possesses interesting biological activities in terms of selective cytotoxicity on cancer cells,

antiproliferative and antimigration effects on endothelial cells. Fisetin and geraldol anticancer and antiangiogenic properties therefore deserve to be optimized in preclinical models, as a single administration or in combination therapy with cytotoxic anticancer agents.

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